

Introduction to Exponential-family Random Graph (ERG or p^*) modeling with *ergm* Version 3.8.0

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1 Getting Started

This vignette is based on the *ergm* tutorial presented at INSNA Sunbelt - St. Pete Beach, Florida, Feb 2011.

Open an R session, and set your working directory to the location where you would like to save this work. You can do this with the pull-down menus (File>Change Dir) or with the command:

```
> setwd('full.path.for.the.folder')
```

To install all of the packages in the statnet suite:

```
> install.packages('statnet')
> library(statnet)
```

Or, to only install the specific statnet packages needed for this tutorial:

```
> install.packages('network')
> install.packages('ergm')
> install.packages('sna')
> library(network)
> library(ergm)
> library(sna)
```

After the first time, to update the packages one can either repeat the commands above, or use:

```
> update.packages('name.of.package')
```

For this tutorial, we will need one additional package (coda), which is recommended (but not required) by ergm:

```
> install.packages('coda')
> library(coda)
```

2 Statistical network modeling; the *ergm* command and *ergm* object

Make sure the statnet package is attached:

```
> library(statnet)
```

or

```
> library(ergm)
> library(sna)
> set.seed(1)
```

The ergm package contains several network data sets that you can use for practice examples.

```
> data(package='ergm') # tells us the datasets in our packages
> data(florentine) # loads flomarriage and flobusiness data
> flomarriage # Let's look at the flomarriage data
```

```
Network attributes:
vertices = 16
directed = FALSE
hyper = FALSE
loops = FALSE
```

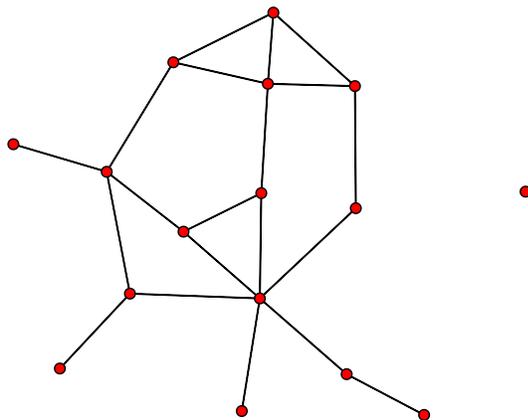
```

multiple = FALSE
bipartite = FALSE
total edges= 20
  missing edges= 0
  non-missing edges= 20

Vertex attribute names:
  priorates totalties vertex.names wealth

No edge attributes
> plot(flomarriage) # Let's view the flomarriage network

```



Remember the general ergm representation of the probability of the observed network, and the conditional log-odds of a tie:

$$\Pr(Y = y) = \exp[\theta'g(y)]/k(\theta)$$

Y is a network; $g(y)$ is a vector of network stats; θ is the vector of coefficients; $k(\theta)$ is a normalizing constant.

$$\text{logit}(\Pr(Y_{ij} = 1|Y^c)) = \theta' \Delta(g(y))_{ij}$$

Y_{ij} is an actor pair in Y ; Y^c is the rest of the network; $\Delta(g(y))_{ij}$ is the change in $g(y)$ when the value of Y_{ij} is toggled on.

We begin with the simplest possible model, the Bernoulli or Erdős-Rényi model, which contains only an edge term.

```

> flomodel.01 <- ergm(flomarriage~edges) # fit model
> flomodel.01

MLE Coefficients:
  edges
-1.609

> summary(flomodel.01) # look in more depth

=====
Summary of model fit
=====

Formula:   flomarriage ~ edges

Iterations: 5 out of 20

Monte Carlo MLE Results:
      Estimate Std. Error MCMC % p-value
edges -1.6094      0.2449      0 <1e-04 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

      Null Deviance: 166.4 on 120 degrees of freedom
      Residual Deviance: 108.1 on 119 degrees of freedom

AIC: 110.1   BIC: 112.9   (Smaller is better.)

```

How to interpret this model? The log-odds of any tie occurring is:

$$\begin{aligned}
 & -1.609 \times \text{change in the number of ties} \\
 & = -1.609 \times 1
 \end{aligned}$$

for all ties, since the addition of any tie to the network changes the number of ties by 1!

Corresponding probability is:

$$\begin{aligned}
 & \exp(-1.609)/(1 + \exp(-1.609)) \\
 & = 0.1667
 \end{aligned}$$

which is what you would expect, since there are 20/120 ties.

Let's add a term often thought to be a measure of "clustering": the number of completed triangles. Note we're in stochastic simulation now – your output will differ

```

> flomodel.02 <- ergm(flomarriage~edges+triangle)
> summary(flomodel.02)

=====
Summary of model fit
=====

Formula:   flomarriage ~ edges + triangle

Iterations: 2 out of 20

```

Monte Carlo MLE Results:

	Estimate	Std. Error	MCMC %	p-value
edges	-1.6709	0.3551	0	<1e-04 ***
triangle	0.1523	0.5924	0	0.798

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Null Deviance: 166.4 on 120 degrees of freedom
Residual Deviance: 108.1 on 118 degrees of freedom

AIC: 112.1 BIC: 117.6 (Smaller is better.)

```
> coef1 = flomodel.02$coef[1]
> coef2 = flomodel.02$coef[2]
> logodds = coef1 + c(0,1,2) * coef2
> expit = function(x) 1/(1+exp(-x))
> ps = expit(logodds)
> coef1 = round(coef1, 3)
> coef2 = round(coef2, 3)
> logodds = round(logodds, 3)
> ps = round(ps, 3)
```

Again, how to interpret coefficients?

Conditional log-odds of two actors forming a tie is:

$-1.671 \times \text{change in the number of ties} + 0.152 \times \text{change in number of triangles}$

- if the tie will not add any triangles to the network, its log-odds is: -1.671 .
- if it will add one triangle to the network, its log-odds is: $-1.671 + 0.152 = -1.519$
- if it will add two triangles to the network, its log-odds is: $-1.671 + 0.152 \times 2 = -1.366$
- the corresponding probabilities are 0.158, 0.18, and 0.203.

Let's take a closer look at the ergm object itself:

```
> class(flomodel.02) # this has the class ergm
[1] "ergm"
> names(flomodel.02) # let's look straight at the ERGM obj.
 [1] "coef"           "sample"         "sample.obs"     "iterations"
 [5] "MCMCtheta"     "loglikelihood"  "gradient"       "hessian"
 [9] "covar"         "failure"        "network"        "newnetworks"
[13] "newnetwork"    "coef.init"      "est.cov"        "coef.hist"
[17] "stats.hist"    "steplen.hist"   "control"        "etamap"
[21] "formula"       "target.stats"   "target.esteq"   "constrained"
[25] "constraints"   "reference"      "estimate"       "offset"
[29] "drop"          "estimable"     "null.lik"       "mle.lik"

> flomodel.02$coef
```

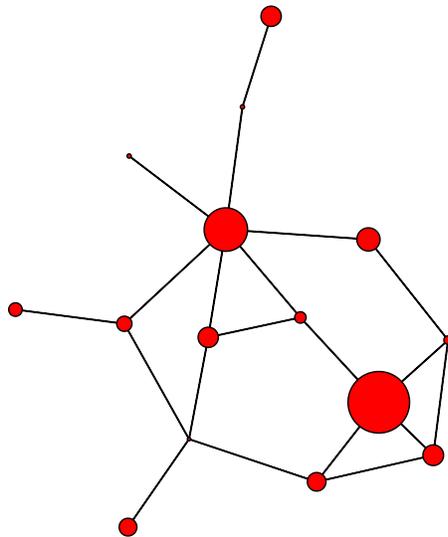
```

      edges  triangle
-1.6709024  0.1522743

> flomodel.02$formula
flomarriage ~ edges + triangle
> flomodel.02$mle.lik
'log Lik.' -54.03178 (df=2)
> wealth <- flomarriage %v% 'wealth' # the %v% extracts vertex
> wealth # attributes from a network

[1] 10 36 55 44 20 32 8 42 103 48 49 3 27 10 146 48
> plot(flomarriage, vertex.cex=wealth/25) # network plot with vertex size
>                                     # proportional to wealth

```



We can test whether edge probabilities are a function of wealth:

```

> flomodel.03 <- ergm(flomarriage~edges+nodecov('wealth'))
> summary(flomodel.03)

=====
Summary of model fit
=====

Formula:  flomarriage ~ edges + nodecov("wealth")

```

Iterations: 4 out of 20

Monte Carlo MLE Results:

	Estimate	Std. Error	MCMC %	p-value
edges	-2.594929	0.536056	0	<1e-04 ***
nodecov.wealth	0.010546	0.004674	0	0.0259 *

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Null Deviance: 166.4 on 120 degrees of freedom

Residual Deviance: 103.1 on 118 degrees of freedom

AIC: 107.1 BIC: 112.7 (Smaller is better.)

Yes, there is a significant positive wealth effect on the probability of a tie.

Let's try a model or two on:

Is there a statistically significant tendency for ties to be reciprocated ('mutuality')?

```
> data(samplk)
> ls() # directed data: Sampson's Monks
> samplk3
> plot(samplk3)
> samplmodel.01 <- ergm(samplk3~edges+mutual)
> summary(samplmodel.01)
```

Let's try a larger network

```
> data(faux.mesa.high)
> mesa <- faux.mesa.high

> plot(mesa)
> mesa
> plot(mesa, vertex.col='Grade')
> legend('bottomleft',fill=7:12,legend=paste('Grade',7:12),cex=0.75)
> fauxmodel.01 <- ergm(mesa ~edges + nodematch('Grade',diff=T) + nodematch('Race',diff=T))
> summary(fauxmodel.01)
```

Note that two of the coefficients are estimated as -Inf (the nodematch coefficients for race Black and Other). Why is this?

```
> table(mesa %v% 'Race') # Frequencies of race
```

```
Black  Hisp  NatAm  Other  White
     6   109    68     4    18
```

```
> mixingmatrix(mesa, "Race")
```

Note: Marginal totals can be misleading
for undirected mixing matrices.

	Black	Hisp	NatAm	Other	White
Black	0	8	13	0	5
Hisp	8	53	41	1	22
NatAm	13	41	46	0	10
Other	0	1	0	0	0
White	5	22	10	0	4

So the problem is that there are very few students in the Black and Other race categories, and these students form no homophilous (within-group) ties. The empty cells are what produce the -Inf estimates.

Time to consider some missing data:

```
> missnet <- network.initialize(10,directed=F)
> missnet[1,2] <- missnet[2,7] <- missnet[3,6] <- 1
> missnet[4,6] <- missnet[4,9] <- NA
> missnet
> plot(missnet)
> ergm(missnet~edges)
```

The coefficient equals -2.590. This is the log-odds of the probability .0698. Our network has 3 ties, out of the 43 nodal pairs (10 choose 2 minus 2) whose dyad status we have observed. $3/43 = 0.0698$.

```
> ergm(missnet~edges+degree(2))
> missnet[4,6] <- missnet[4,9] <- 0
> ergm(missnet~edges+degree(2))
```

The two estimates for the degree 2 coefficient differ considerably. In the first case, there is one node we know for sure has degree 2, two that may or may not, and seven that we know for sure do not. In the latter, there is one node that has degree 2, and nine that do not.

3 Model terms available for *ergm* estimation and simulation

Model terms are the expressions (e.g. “triangle”) used to represent predictors on the right-hand side of equations used in:

- calls to `ergm` (to estimate an ergm model)
- calls to `simulate` (to simulate networks from an ergm model fit)
- calls to `summary` (to obtain measurements of network statistics on a dataset)

3.1 Terms provided with `ergm`

For a list of available terms that can be used to specify an ERGM, see Appendix B, or type:

```
help('ergm-terms')
```

For a more complete discussion of these terms see the ‘Specifications’ paper in J Stat Software v. 24. (link is available online at www.statnet.org)

3.2 Coding new terms

We have recently released a new package (`ergm.userterms`) and tutorial aimed at making it much easier than before to write one's own terms. The package is available on CRAN, and installing it will also download the tutorial (`ergmuserterms.pdf`). We teach a workshop at the Sunbelt meetings, and are also hoping for the tutorial to appear soon in the *Journal of Statistical Software*. Note that writing up new `ergm` terms requires some knowledge of C and the ability to build R from source (although the latter is covered in the tutorial).

4 Network simulation: the *simulate* command and *network.list* objects

Once we have estimated the coefficients of an ERGM, the model is completely specified. It defines a probability distribution across all networks of this size. If the model is a good fit to the observed data, then networks drawn from this distribution will be more likely to "resemble" the observed data. To see examples of networks drawn from this distribution we use the `simulate` command:

```
> flomodel.03.sim <- simulate(flomodel.03,nsim=10)
> class(flomodel.03.sim)
```

```
[1] "network.list"
```

```
> summary(flomodel.03.sim)
```

```
Number of Networks: 10
Model: flomarriage ~ edges + nodecov("wealth")
Reference: ~Bernoulli
Constraints: ~.
Parameters:
      edges nodecov.wealth
-2.59492903    0.01054591
```

```
Stored network statistics:
```

```
      edges nodecov.wealth
[1,]    24          2736
[2,]    21          2389
[3,]    20          2203
[4,]    25          2803
[5,]    26          2835
[6,]    22          2428
[7,]    21          1965
[8,]    20          2161
[9,]    21          1977
[10,]   22          2293
```

```
> length(flomodel.03.sim)
```

```
[1] 10
```

```
> flomodel.03.sim[[1]]
```

```
Network attributes:
  vertices = 16
  directed = FALSE
```

```

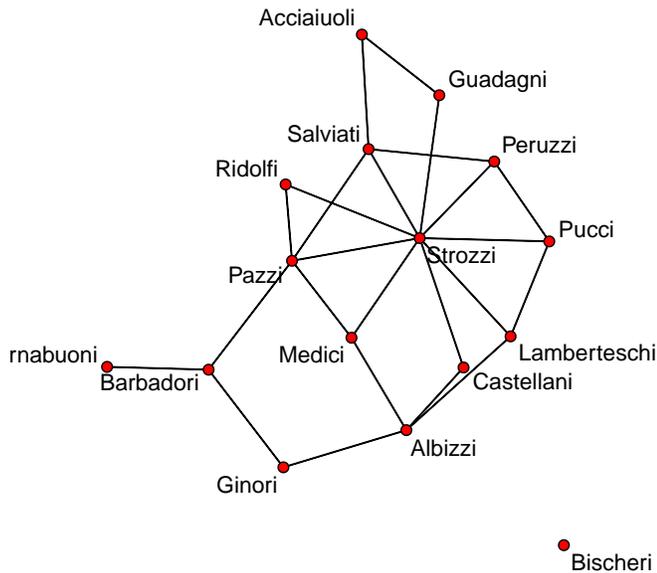
hyper = FALSE
loops = FALSE
multiple = FALSE
bipartite = FALSE
total edges= 24
  missing edges= 0
  non-missing edges= 24

Vertex attribute names:
  priorates totalties vertex.names wealth

No edge attributes

> plot(flomodel.03.sim[[1]], label= flomodel.03.sim[[1]] %% "vertex.names")

```



Voila. Of course, yours will look somewhat different.

5 Examining the quality of model fit – *GOF*

ERGMs are generative models – that is, they represent the process that governs tie formation at a local level. These local processes in turn aggregate up to produce characteristic global network properties, even though these global properties are not explicit terms in the model. One test of whether a model “fits the data” is therefore how well it reproduces these global properties. We do this by choosing a network statistic that is not in the model, and comparing the value of this statistic observed in the original network to the distribution of values we get in simulated networks from our model.

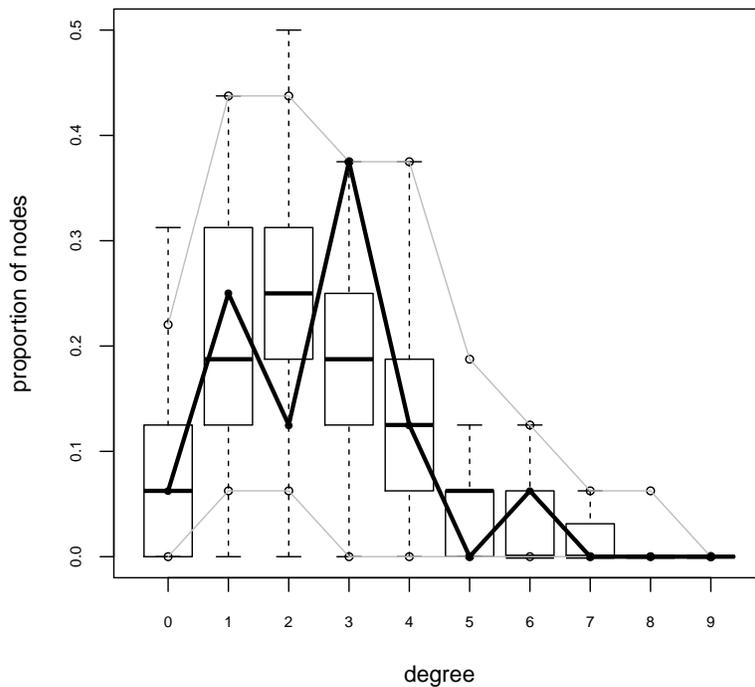
```
> flomodel.03.gof <- gof(flomodel.03~degree)
> flomodel.03.gof
```

Goodness-of-fit for degree

	obs	min	mean	max	MC	p-value
0	1	0	1.23	5		1.00
1	4	0	3.58	7		0.92
2	2	0	4.16	8		0.30
3	6	0	3.10	8		0.22
4	2	0	2.10	7		1.00
5	0	0	0.91	3		0.76
6	1	0	0.53	2		0.94
7	0	0	0.27	3		1.00
8	0	0	0.09	2		1.00
9	0	0	0.02	1		1.00
10	0	0	0.01	1		1.00

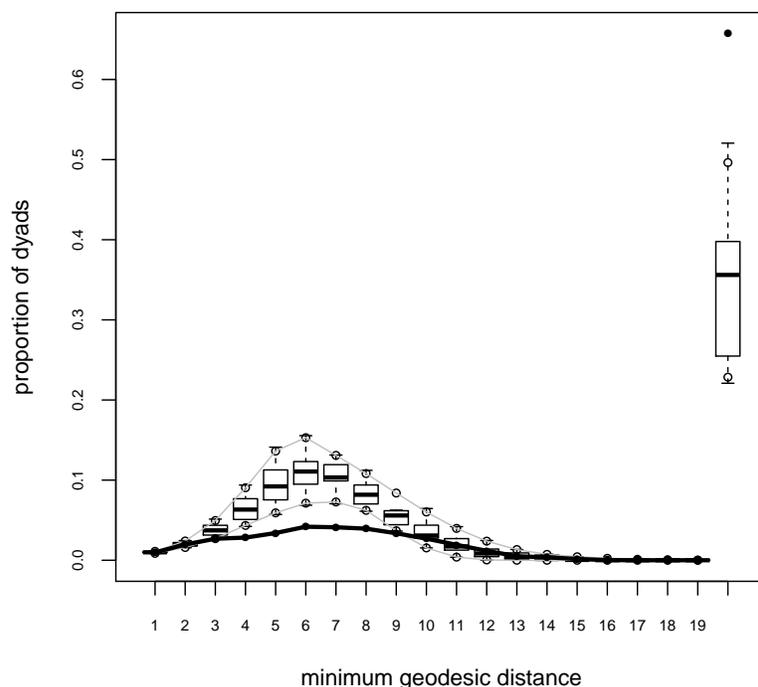
```
> plot(flomodel.03.gof)
```

Goodness-of-fit diagnostics



```
> mesamodel.02 <- ergm(mesa~edges)
> mesamodel.02.gof <- gof(mesamodel.02~distance,control=control.gof.formula(nsim=10))
> plot(mesamodel.02.gof)
>
```

Goodness-of-fit diagnostics



For a good example of model exploration and fitting for the Add Health Friendship networks, see Goodreau, Kitts & Morris, *Demography* 2009.

6 Diagnostics: troubleshooting and checking for model degeneracy

The computational algorithms in `ergm` use MCMC to estimate the likelihood function. Part of this process involves simulating a set of networks to approximate unknown components of the likelihood.

When a model is not a good representation of the observed network the estimation process may be affected. In the worst case scenario, the simulated networks will be so different from the observed network that the algorithm fails altogether. This can occur for two general reasons. First, the simulation algorithm may fail to converge, and the sampled networks are thus not from the specified distribution. Second, the model parameters used to simulate the networks are too different from the MLE, so even though the simulation algorithm is producing a representative sample of networks, this is not the sample that would be produced under the MLE.

For more detailed discussions of model degeneracy in the ERGM context, see the papers in *J Stat Software* v. 24. (link is available online at www.statnet.org)

We can use diagnostics to see what is happening with the simulation algorithm, and these can lead us to ways to improve it.

We will first consider a simulation where the algorithm works. To understand the algorithm, consider

```
> fit <- ergm(flobusiness~edges+degree(1),  
+ control=control.ergm(MCMC.interval=1, MCMC.burnin=1000, seed=1))
```

This runs a version with every network returned. Let us look at the diagnostics produced:

```
> mcmc.diagnostics(fit, center=F)
```

Let's look more carefully at a default model fit:

```
> fit <- ergm(flobusiness~edges+degree(1))
> mcmc.diagnostics(fit, center=F)
```

Now let us look at a more interesting case, using a larger network:

```
> data('faux.magnolia.high')
> magnolia <- faux.magnolia.high
> plot(magnolia, vertex.cex=.5)

> fit <- ergm(magnolia~edges+triangle, control=control.ergm(seed=1))
> mcmc.diagnostics(fit, center=F)
```

```
Error in ergm.MCMLE(init, nw, model, initialfit = (initialfit <- NULL), :
  Number of edges in a simulated network exceeds that in the observed by a factor of more than 20. Th
```

Very interesting. This model produced degenerate networks. You could have gotten some more feedback about this during the fitting, by using:

```
> fit <- ergm(magnolia~edges+triangle, control=control.ergm(seed=1), verbose=T)
```

You might try to increase the MCMC sample size:

```
> fit <- ergm(magnolia~edges+triangle,seed=1,
+ control = control.ergm(seed=1, MCMC.samplesize=20000),
+ verbose=T)
> mcmc.diagnostics(fit, center=F)
```

How about trying the more robust version of modeling triangles: GWESP? (For a technical introduction to GWESP see Hunter and Handcock; for a more intuitive description and empirical application see Goodreau Kitts and Morris 2009)

```
> fit <- ergm(magnolia~edges+gwap(0.5, fixed=T),
+ control = control.ergm(seed=1))
> mcmc.diagnostics(fit)
```

Still degenerate, but maybe getting closer?

```
> fit <- ergm(magnolia~edges+gwap(0.5, fixed=T)+nodematch('Grade')+nodematch('Race')+
+ nodematch('Sex'),
+ control = control.ergm(seed=1),
+ verbose=T)
> pdf('diagnostics1.pdf') #Use the recording function if possible, otherwise send to pdf
> mcmc.diagnostics(fit)
> dev.off() #If you saved to pdf, look at the file
> fit <- ergm(magnolia~edges+gwap(0.25, fixed=T)+nodematch('Grade')+nodematch('Race')+
+ nodematch('Sex'),
+ control = control.ergm(seed=1))
> mcmc.diagnostics(fit)
>
```

One more try...

```
> fit <- ergm(magnolia~edges+gwesp(0.25,fixed=T)+nodematch('Grade')+nodematch('Race')+
+   nodematch('Sex'),
+   control = control.ergm(seed=1,MCMC.samplesize=4096,MCMC.interval=8192),
+   verbose=T)
```

```
> mcmc.diagnostics(fit)
```

Sample statistics summary:

Iterations = 131072:134340608
Thinning interval = 8192
Number of chains = 1
Sample size per chain = 16384

1. Empirical mean and standard deviation for each variable,
plus standard error of the mean:

	Mean	SD	Naive SE	Time-series SE
edges	2.441	46.40	0.3625	3.689
gwesp.fixed.0.25	3.143	41.93	0.3275	4.069
nodematch.Grade	2.880	44.11	0.3446	3.810
nodematch.Race	3.618	42.73	0.3338	3.935
nodematch.Sex	1.972	37.52	0.2931	3.197

2. Quantiles for each variable:

	2.5%	25%	50%	75%	97.5%
edges	-84.00	-29.00	1.00	33.00	99.00
gwesp.fixed.0.25	-72.71	-25.47	1.13	29.68	94.51
nodematch.Grade	-79.00	-27.00	2.00	31.00	96.00
nodematch.Race	-75.00	-26.00	1.00	31.00	93.00
nodematch.Sex	-68.00	-24.00	1.00	27.00	78.00

Sample statistics cross-correlations:

	edges	gwesp.fixed.0.25	nodematch.Grade	nodematch.Race
edges	1.0000000	0.8497712	0.9607215	0.9460052
gwesp.fixed.0.25	0.8497712	1.0000000	0.8688021	0.8483614
nodematch.Grade	0.9607215	0.8688021	1.0000000	0.9183533
nodematch.Race	0.9460052	0.8483614	0.9183533	1.0000000
nodematch.Sex	0.9050235	0.7930151	0.8739491	0.8583987
	nodematch.Sex			
edges	0.9050235			
gwesp.fixed.0.25	0.7930151			
nodematch.Grade	0.8739491			
nodematch.Race	0.8583987			
nodematch.Sex	1.0000000			

Sample statistics auto-correlation:

Chain 1

	edges	gwest.fixed.0.25	nodematch.Grade	nodematch.Race
Lag 0	1.0000000	1.0000000	1.0000000	1.0000000
Lag 8192	0.7798096	0.9818971	0.8302156	0.8175705
Lag 16384	0.7324306	0.9655592	0.7830802	0.7693207
Lag 24576	0.7050110	0.9502349	0.7577113	0.7435499
Lag 32768	0.6896580	0.9361270	0.7419942	0.7289507
Lag 40960	0.6728069	0.9227838	0.7266955	0.7141575

	nodematch.Sex
Lag 0	1.0000000
Lag 8192	0.7820718
Lag 16384	0.7275206
Lag 24576	0.6979356
Lag 32768	0.6793173
Lag 40960	0.6658113

Sample statistics burn-in diagnostic (Geweke):
Chain 1

Fraction in 1st window = 0.1
Fraction in 2nd window = 0.5

	edges	gwest.fixed.0.25	nodematch.Grade	nodematch.Race
	0.4202	0.3199	0.4055	0.4147

	nodematch.Sex
	0.7801

Individual P-values (lower = worse):

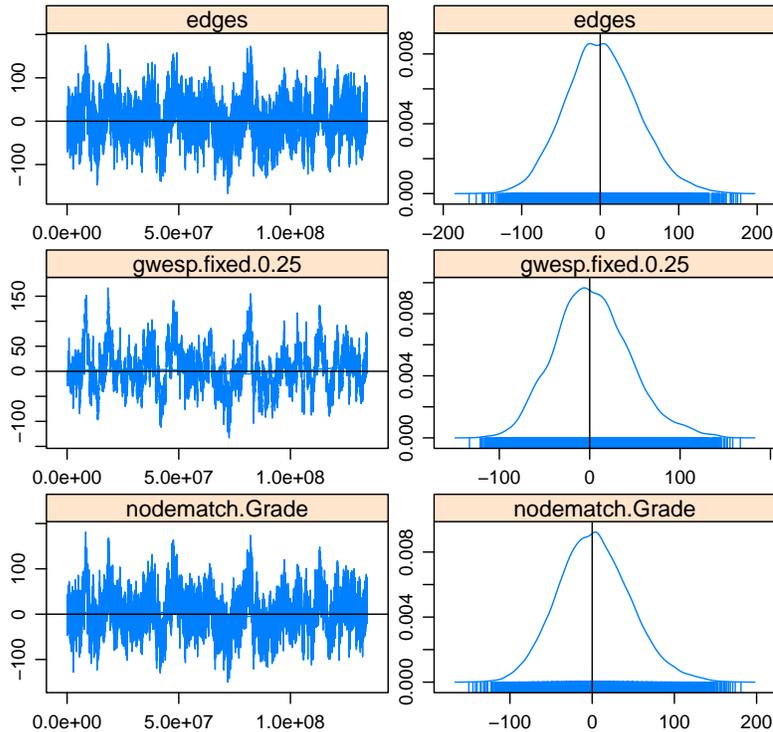
	edges	gwest.fixed.0.25	nodematch.Grade	nodematch.Race
	0.6743736	0.7490473	0.6851239	0.6783401

	nodematch.Sex
	0.4353459

Joint P-value (lower = worse): 0.5131558 .

MCMC diagnostics shown here are from the last round of simulation, prior to computation of final para

Sample statistics



Success! Of course, in real life one might have a lot more trial and error.

Changes in version 3.2 of the `ergm` estimation algorithm mean that the MCMC diagnostic plots can no longer be used to ensure that the mean statistics from the model match the observed network statistics. For that functionality, please use the GOF command: `gof(fit, GOF=~model)`. The plots can still be used to assess MCMC mixing and convergence.

7 Working with egocentrically sampled network data

In many empirical contexts, it is not feasible to collect a network census or even an adaptive (link-traced) sample. Even when one of these may be possible in practice, egocentrically sampled data are typically cheaper and easier to collect.

Long regarded as the poor country cousin in the network data family, egocentric data contain a remarkable amount of information. With the right statistical methods, such data can be used to explore the properties of the complete networks in which they are embedded. The basic idea here is to combine what is observed, with assumptions, to define a class of models that describe the distribution of networks that are centered on the observed properties. The variation in these networks quantifies some of the uncertainty introduced by the assumptions.

Let's start with a simple fictional example: You have a sample of persons who were asked about the friends they had seen face-to-face more than once in the last week. The respondent was asked their own sex, and the sex of each friend (for up to three friends). Summary statistics from these data thus include the sex distribution, the degree distribution (it could be broken down by sex, but we will just examine the marginal distribution here), and the joint distribution of the respondent and friend's sex (the sex mixing matrix). Let's assume there are equal numbers of men and women in the sampled respondents. The other distributions are shown below:

Degree distribution				Sex mixing matrix		
Degree	Frequency	Fraction	Ties	(410 total)	Friend	
0	180	0.36	0		M	F
1	245	0.49	245			
2	60	0.12	120	Respondent	M	F
3	15	0.03	45		F	
Total	500	1.00	410			

So, total N respondents = 500, total N friends reported = 410.

We can use an ERGM to fit the parameters associated with these observed statistics, then use the fitted model to simulate complete networks that are drawn from the distribution of networks that is centered around these statistics. As a theoretical exercise, this provides a method for investigating the complete network implications of these observed summary statistics. As an empirical exercise, the primary assumption needed for inference is that the data we have is sampled from a population in equilibrium (and, as in all statistical inference, that our model is correct). The theory for this is developed in Krivitsky, 2009.

We need to make assumptions about size, directedness and bipartite-ness when we model and simulate the complete network.

- **Size:** any size can be simulated, but if the model is fit using the observed frequencies, it should be used to simulate a population of that size, unless a size adjustment is made in the simulation (see Krivitsky, Handcock and Morris 2011). We are going to work with a population size 500 here, equal to the number of respondents.
- **Directedness:** Ego data are in one sense inherently directed (respondents nominate alters, alters are not observed), but the relationship may be either. In this case (“seen more than once”) it is undirected, so we will fit and simulate an undirected network.
- **Bipartite:** Ego data can be bipartite (if no alters are also respondents, or data are collected on 2-mode networks) or not (if respondents are also alters). But again, the relationship may be either. “Seen” is undirected, and we will fit and simulated and undirected network.

In sum, we will simulate a one-mode, undirected network of size 500, assuming the ego statistics we observed contain the information we need to calculate the statistics that would have been observed in a self-contained population of that size, noting that other assumptions are possible.

To ensure consistency between the degree distribution (which is a tabulation of nodes) and the mixing matrix (which is a cross-tabulation of ties) in our simulated “complete network,” it is important to recognize that in a complete network, the degree distribution should imply twice the number of ties observed in the mixing matrix, because every tie is being reported by both nodes in the degree distribution. If we are fixing the population size at 500 in this simulation, then our observed mixing matrix data needs to be divided by 2.

Start by initializing an empty network of the desired size and assign the “sex” attribute to the nodes:

```
> ego.net <- network.initialize(500, directed=F)
> ego.net %v% 'sex' <- c(rep(0,250),rep(1,250))
```

Set up the observed statistics (adjusted for this “complete” network) as we will use them to assess the accuracy of the simulation later:

```
> ego.deg <- c(180, 245, 60, 15) # node distn
> ego.mixmat <- matrix(c(164,44,26,176)/2, nrow=2, byrow=T) # adjusted tie distn
```

Then, pick the observed statistics you want to target – we will start with a simple model here: the total number of ties (edges), and the number of sex-matched ties (homophily). These are both functions of the observed statistics:

```
> ego.edges <- sum(ego.mixmat)
> ego.sexmatch <- ego.mixmat[1,1]+ego.mixmat[2,2]
```

And combine these into a vector

```
> ego.target.stats <- c(ego.edges, ego.sexmatch)
> ego.target.stats

[1] 205 170
```

Now, fit an ERGM to this “network”, with terms for the statistics you want to match, and the “target.stats” argument for **ergm** that specifies the target values for those statistics:

```
> ego.fit <- ergm(ego.net ~ edges + nodematch('sex'),
+ target.stats = ego.target.stats)
```

Take a look at the fitted model:

```
> summary(ego.fit)

=====
Summary of model fit
=====

Formula:   nw ~ edges + nodematch("sex")
<environment: 0x55ef8ec11688>

Iterations: 8 out of 20

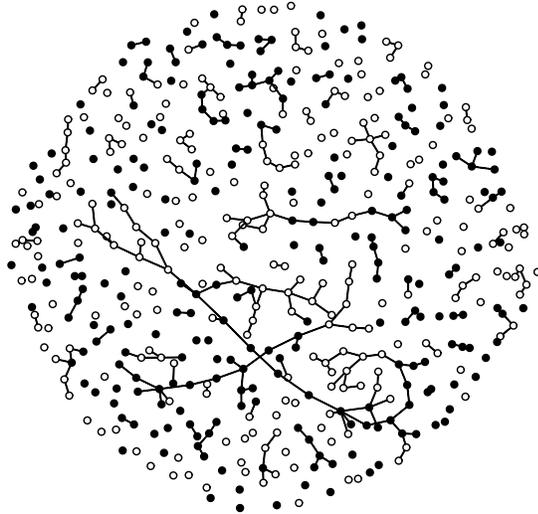
Monte Carlo MLE Results:
              Estimate Std. Error MCMC % p-value
edges          -7.4870    0.1690     0 <1e-04 ***
nodematch.sex   1.5866    0.1857     0 <1e-04 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Null Deviance: 172940 on 124750 degrees of freedom
Residual Deviance: 2941 on 124748 degrees of freedom

AIC: 2945    BIC: 2964    (Smaller is better.)
```

Now that you have a fitted model, you can simulate a complete network from it, and look at the results:

```
> ego.sim1 <- simulate(ego.fit)
> plot(ego.sim1, vertex.cex=.65, vertex.col="sex")
```



Does it reproduce the observed degree and mixing frequencies? We only targeted the total number of edges, not the full degree distribution.

```
> rbind(summary(ego.sim1 ~ degree(c(0:3))), ego.deg)
```

	degree0	degree1	degree2	degree3
ego.deg	206	181	77	28

```
> mixingmatrix(ego.sim1, "sex")
```

Note: Marginal totals can be misleading for undirected mixing matrices.

	0	1
0	91	42
1	42	94

```
> ego.mixmat
```

	[,1]	[,2]
[1,]	82	22
[2,]	13	88

We only targeted the number of same-sex ties, not the full mixing matrix.

The simulation stats seem quite different than the observed stats, and there are two possible reasons: either we mis-specified the original model (bias), or this one random draw may be unrepresentative of the distribution described by the model (variance). The latter is easily examined by simulating 100 networks, to see where the observed data fall in the distribution of networks produced by the model:

```
> ego.sim100 <- simulate(ego.fit, nsim=100)
> ego.sim100
```

```
Number of Networks: 100
Model: nw ~ edges + nodematch("sex")
Reference: ~Bernoulli
Constraints: ~.
Parameters:
      edges nodematch.sex
-7.487013      1.586633
```

More information can be obtained with

```
> summary(ego.sim100)
```

First, we'll look at how well the simulations reproduced the target statistics that were included in the model (note, not the observed full distributions):

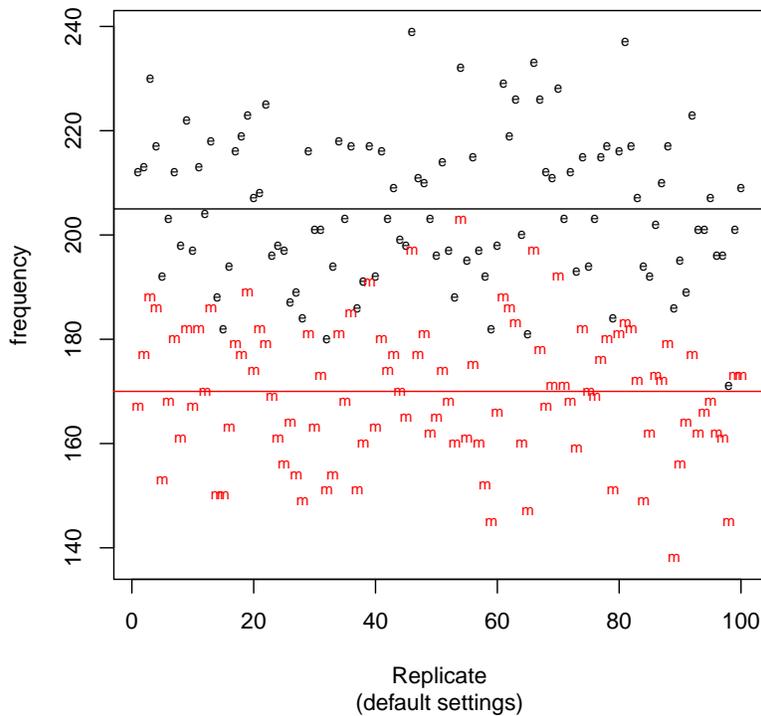
```
> sim.stats <- attr(ego.sim100,"stats")
> rbind(colMeans(sim.stats), ego.target.stats)
```

```
              edges nodematch.sex
ego.target.stats 205.22      169.89
ego.target.stats 205.00      170.00
```

These look pretty good – the means of the simulated target stats are within 1% of the observed. We can plot the 100 replicates to see check the variation for any problematic patterns:

```
> matplot(1:nrow(sim.stats), sim.stats,
+   pch=c("e","m","0","+"), cex=.65,
+   main="100 simulations from ego.fit model", sub="(default settings)",
+   xlab="Replicate", ylab="frequency")
> abline(h=ego.target.stats, col=c(1:4))
```

100 simulations from ego.fit model



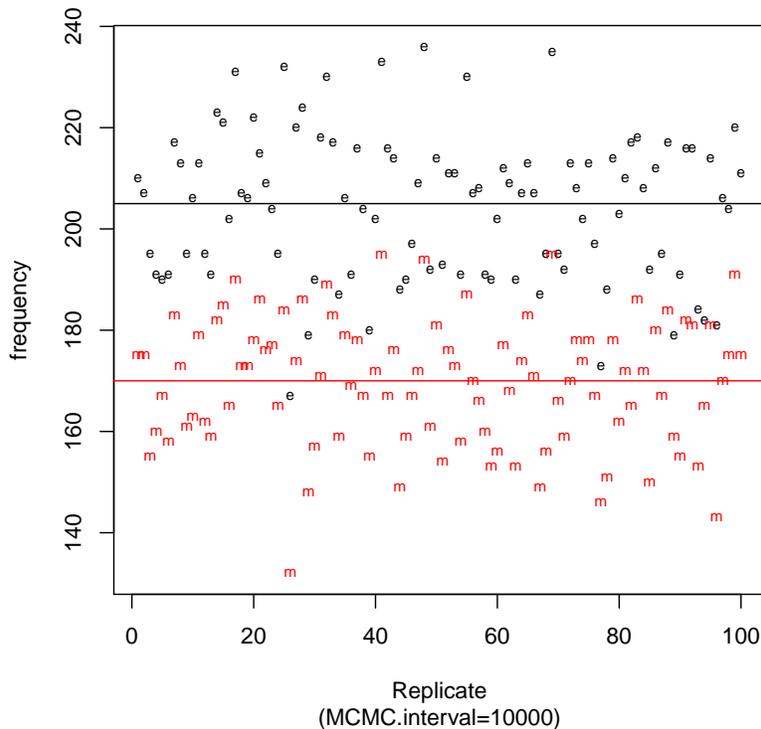
The lines mark the target statistic frequencies in the observed data. The points represent the frequencies in the simulated networks.

The simulated network statistics vary stochastically around the target values, not trending over time.

But, there is clear autocorrelation across the replicates, which suggests we might want to increase the MCMC interval to draw more independent realizations.

```
> ego.sim100 <- simulate(ego.fit, nsim=100,  
+   control=control.simulate.ergm(MCMC.interval=10000))  
> sim.stats <- attr(ego.sim100,"stats")  
> matplot(1:nrow(sim.stats), sim.stats,  
+   pch=c("e","m"), cex=.65,  
+   main="100 simulations from ego.fit model", sub="(MCMC.interval=10000)",  
+   xlab="Replicate", ylab="frequency")  
> abline(h=ego.target.stats, col=c(1:2))
```

100 simulations from ego.fit model



With the larger interval, the autocorrelation is no longer detectable, and all of the statistics from the simulated networks vary in a symmetric band around their targets.

The variation (about $\pm 10\%$) represents the range of target statistics that are consistent with the fitted coefficients.

If you wanted instead to constrain these statistics to equal a specified value, then you can use the "constraints=" argument during the `ergm` fit instead.

This is good for verifying that the simulation reproduces the target values we specified. So now let's see whether the simulated complete networks also match statistics that were not set by the targets. We targeted edges and homophily. How well does this model reproduce the full degree distribution?

```
> sim.fulldeg <- summary(ego.sim100 ~ degree(c(0:10)))
> colnames(sim.fulldeg) <- paste("deg",0:10, sep='')
> sim.fulldeg[1:5,]
      deg0 deg1 deg2 deg3 deg4 deg5 deg6 deg7 deg8 deg9 deg10
[1,]  211  188   75   22   4   0   0   0   0   0   0
[2,]  216  184   74   24   1   0   1   0   0   0   0
[3,]  228  180   72   16   2   2   0   0   0   0   0
[4,]  245  163   66   20   4   1   1   0   0   0   0
[5,]  227  190   63   16   4   0   0   0   0   0   0
```

Recall that the degree range in our data was $[0,3]$ by design, but we did not constrain the simulations to this range. If our model correctly captured the processes that led to the aggregate statistics we observe in our data, the simulated networks would show us what we missed. Here the simulated networks suggest that the fully observed network would have a wider range of degrees, which we might

have observed, had we not truncated the data collection at 3 friends per respondent. About 1% of nodes have degree 4 or 5, and the max observed is 6.

But did our model did correctly capture the underlying processes? How well does the simulated degree distribution from this model match the frequencies we did observe? Aggregating the degrees of 3 or more in the simulations, we find:

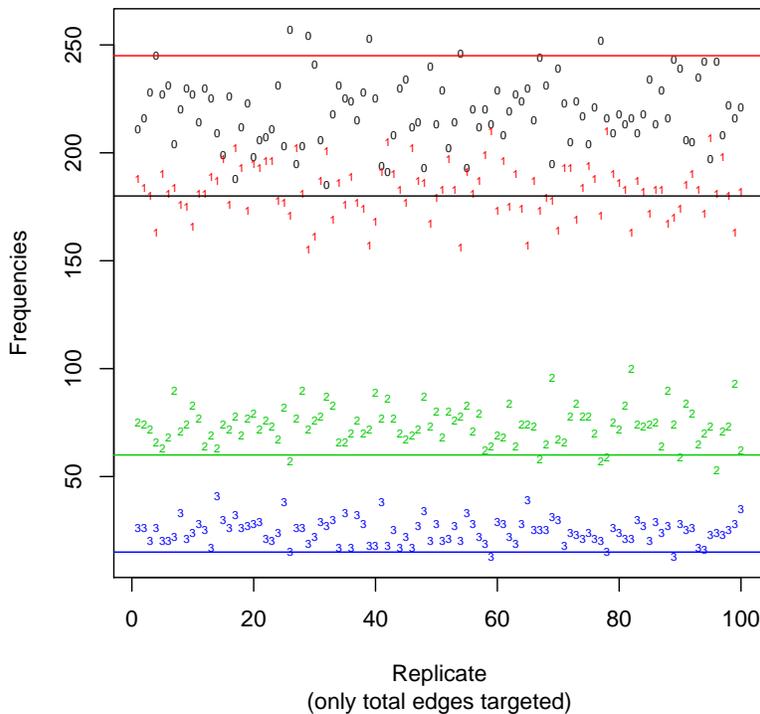
```
> sim.deg <- cbind(sim.fulldeg[,1:3], apply(sim.fulldeg[,4:11],1,sum))
> colnames(sim.deg) <- c(colnames(sim.fulldeg)[1:3],"degree3+")
> rbind(colMeans(sim.deg),ego.deg)

      deg0  deg1  deg2  degree3+
ego.deg 180.00 245.00 60.00   15.00
```

As with the single simulation above, the discrepancies between the simulation averages and the observed statistics are quite large. We can see this more clearly by plotting the degree frequencies for the 100 replicate networks we simulated:

```
> matplot(1:nrow(sim.deg), sim.deg, pch=as.character(0:3), cex=.5,
+ main="Comparing ego.sims to non-targeted degree frequencies",
+ sub = "(only total edges targeted)",
+ xlab = "Replicate", ylab = "Frequencies")
> abline(h=c(180, 245, 60, 15), col=c(1:4))
```

Comparing ego.sims to non-targeted degree frequencies



The simulations are producing systematically more isolates than expected (the “0” points vs. the black line), and systematically more degree 1 nodes. In fact, the two degree frequencies are essentially reversed in the simulation.

The fraction of nodes with 2 or 3+ partners is systematically off but by a much smaller amount.

So our observed network has fewer isolates than expected in a network of this density, more degree 1 nodes than expected, and fewer degree 2 and 3+ nodes.

This suggests the model is mis-specified. Since the degree 0 vs. degree 1 is the worst fitting aspect, we will try using the number of isolates as a target statistic in the model.

```
> ego.isolates <- ego.deg[1]
> ego.target.stats <- c(ego.edges, ego.sexmatch, ego.isolates)
> ego.fit <- ergm(ego.net ~ edges + nodematch('sex') + degree(0),
+ target.stats = ego.target.stats)
> summary(ego.fit)
```

```
=====
Summary of model fit
=====
```

```
Formula: nw ~ edges + nodematch("sex") + degree(0)
<environment: 0x55ef91e8fb58>
```

```
Iterations: 2 out of 20
```

```
Monte Carlo MLE Results:
```

	Estimate	Std. Error	MCMC %	p-value
edges	-8.3939	0.2500	0	<1e-04 ***
nodematch.sex	1.5761	0.1860	0	<1e-04 ***
degree0	-0.9602	0.1634	0	<1e-04 ***

```
---
```

```
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
Null Deviance: 172940 on 124750 degrees of freedom
Residual Deviance: 2903 on 124747 degrees of freedom
```

```
AIC: 2909 BIC: 2939 (Smaller is better.)
```

Simulating from this model, the target statistics are again well matched:

```
> ego.sim100 <- simulate(ego.fit, nsim=100,
+ control=control.simulate.ergm(MCMC.interval=10000))
> sim.stats <- attr(ego.sim100,"stats")
> rbind(colMeans(sim.stats), ego.target.stats)
```

	edges	nodematch.sex	degree0
ego.target.stats	204.43	169.89	180.78
ego.sim100	205.00	170.00	180.00

And the full degree frequencies look much better:

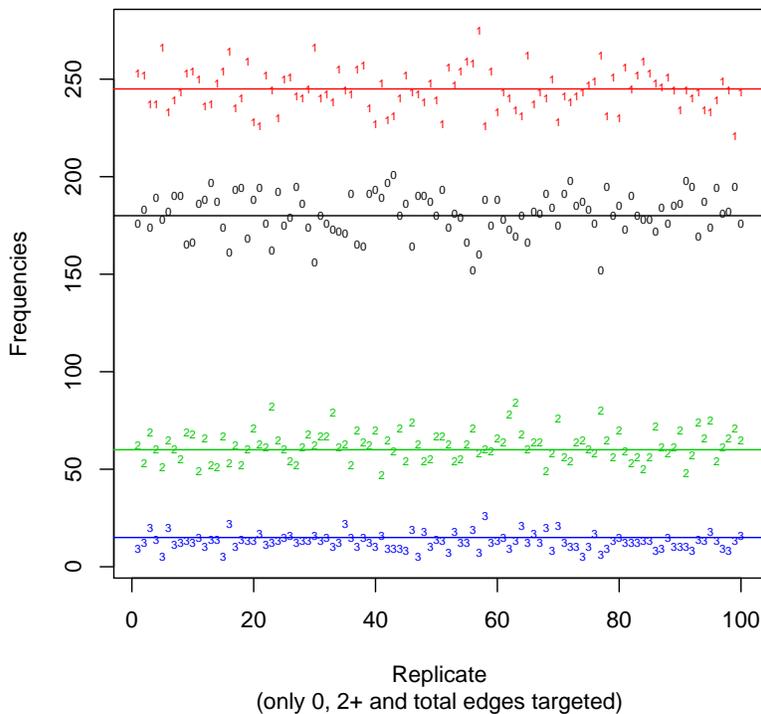
```
> sim.fulldeg <- summary(ego.sim100 ~ degree(c(0:10)))
> sim.deg <- cbind(sim.fulldeg[,1:3], apply(sim.fulldeg[,4:11],1,sum))
> colnames(sim.deg) <- c(colnames(sim.fulldeg)[1:3],"degree3+")
> rbind(colMeans(sim.deg),ego.deg)
```

	degree0	degree1	degree2	degree3+
	180.78	244.16	62.29	12.77
ego.deg	180.00	245.00	60.00	15.00

and finally, plotting the full degree frequencies

```
> matplot(1:nrow(sim.deg), sim.deg, pch=as.character(0:3), cex=.5,
+ main="Comparing ego.sims to non-targeted degree frequencies",
+ sub = "(only 0, 2+ and total edges targeted)",
+ xlab = "Replicate", ylab = "Frequencies")
> abline(h=c(180, 245, 60, 15), col=c(1:4))
```

Comparing ego.sims to non-targeted degree frequencies



The degree frequencies in these simulated networks are now well centered on the observed frequencies. So adding the one additional parameter to capture the lower than expected number of isolates did a good job of capturing how our observed network deviates from a random network with this density.

The fraction of nodes with 3+ partners produced by our new model might still be a bit low.

Moral: We can use ERGMs to estimate network models using target statistics from egocentrically sampled data. The fact that the target statistics are reproduced by this model does not guarantee that additional features of the network would also be reproduced. But starting with simple models can help to identify whether and how the aggregate statistics we observe from an egocentric sample deviate from those we would expect from the model. If we fit all of the observed statistics without a saturated model, we cannot reject the hypothesis that this model produced the network we sampled from.

We can also use this approach to explore network statistics that are not visible at all from the egocentric data, e.g., the geodesic distribution, betweenness, etc., but it must always be remembered

that the distributions we will produce are based on our model. They faithfully reproduce the model, but that does not mean that the model faithfully represents the population.

In the STERGM workshop, we show how complete dynamic networks also can be simulated over time on the basis of egocentric data like these, with the minimal addition of a single estimate of partnership duration. For a movie of an example dynamic simulation used to explore the impact of network structure on HIV transmission, see statnet.org/movies.

8 Additional functionality in the statnet family of packages

8.1 In the `ergm` and `network` packages:

- ERGMs for valued ties – see the paper by Pavel Krivitsky (2012)
- Analysis of bipartite networks – `network` recognizes this as an attribute of the network and `ergm` provides specific model terms for such networks that begin with `b1` or `b2` (try: `search.ergmTerms(categories=c('bipartite'))`).

8.2 In other packages from the statnet suite:

These are in stand-alone packages that can be downloaded either from CRAN, or from the `statnet` website. Many have online training materials from our workshops. For more detailed information, please visit the `statnet` webpage (<http://statnet.org>).

8.2.1 Static (cross-sectional) network analysis packages

- `sna` – Traditional SNA methods and summaries.
- `latentnet` – Latent space and latent cluster analysis.
- `netperm` – Network permutation models.
- `degreenet` – MLE estimation for degree distributions (negative binomial, Poisson, scale-free, etc.)
- `networksis` – Simulation of bipartite networks with given degree distributions.

8.2.2 Dynamic (longitudinal) network analysis packages:

- `tergm` – Temporal ERGMs (TERGMs): discrete-time dynamic network models for longitudinal network panel data, and other temporal extensions.
- `relevent` – Relational event models: continuous-time dynamic network models for longitudinal network data.
- `networkDynamic` – Dynamic network data storage and manipulation.
- `ndtv` – Network movie maker.

8.3 R packages that build on statnet

There is a growing number of R packages written by other folks that build on or extend the functionality of the statnet suite. You can get a current list of those packages by looking at the reverse depends/suggests on CRAN. A partial list includes:

- **EpiModel** package – Mathematical Modeling of Infectious Disease, includes functions for deterministic compartmental modeling, stochastic individual contact modeling, and stochastic network modeling
- **RDS** package – Estimation with data collected using Respondent-Driven Sampling.
- **Bergm** package – Bayesian ERGM estimation
- **hergm** package – Hierarchical Exponential-Family Random Graph Models with Local Dependence (for latent groups).
- **lvm4net** package – Latent variable models.
- **VBLPCM** package – Variational Bayes Latent Position Cluster Models.
- **xergm** package – Temporal exponential random graph models (TERGM) by bootstrapped pseudolikelihood, MCMC MLE and (temporal) network autocorrelation models.

8.4 Additional functionality in development:

- **ergm.ego** package – ERGM estimation and inference from egocentrically sampled data (expected May 2015)
- **tsna** package – Temporally extended (vertex and edge) SNA methods for dynamic longitudinal network data (expected May 2015)
- **MLergm** package – ERGM estimation and inference for multi-level data (for observed groups) (expected 2016)

9 Statnet Commons: The core development team

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Appendix A: Clarifying the terms – ergm and network

You will see the terms `ergm` and `network` used in multiple contexts throughout the documentation. This is common in R, but often confusing to newcomers. To clarify:

`ergm`

- **ERGM**: the acronym for an Exponential Random Graph Model; a statistical model for relational data that takes a generalized exponential family form.

- **ergm package**: one of the packages within the **statnet** suite
- **ergm function**: a function within the **ergm** package; fits an ERGM to a network object, creating an **ergm** object in the process.
- **ergm object**: a class of objects produced by a call to the **ergm** function, representing the results of an ERGM fit to a network.

network

- **network**: a set of actors and the relations among them. Used interchangeably with the term graph.
- **network package**: one of the packages within the **statnet** suite; used to create, store, modify and plot the information found in network objects.
- **network object**: a class of object in R used to represent a network.

References

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